1. FFF Techuologies, FFF-Scaffold Mnuf. 2. Scaffold Manuf. Technologies 3. Non-degradation Scaffold 4.BONE Tissue Eng. Scaffolds 5. 3-D cell Assembled 6. Laser Directed Guided Writing(LDGW) of cell

Prof. Yongnian Yan

BONE Tissue Scaffolds (Degradable)



Scaffold poly (L-lactic acid) Tricalcium Phosphate



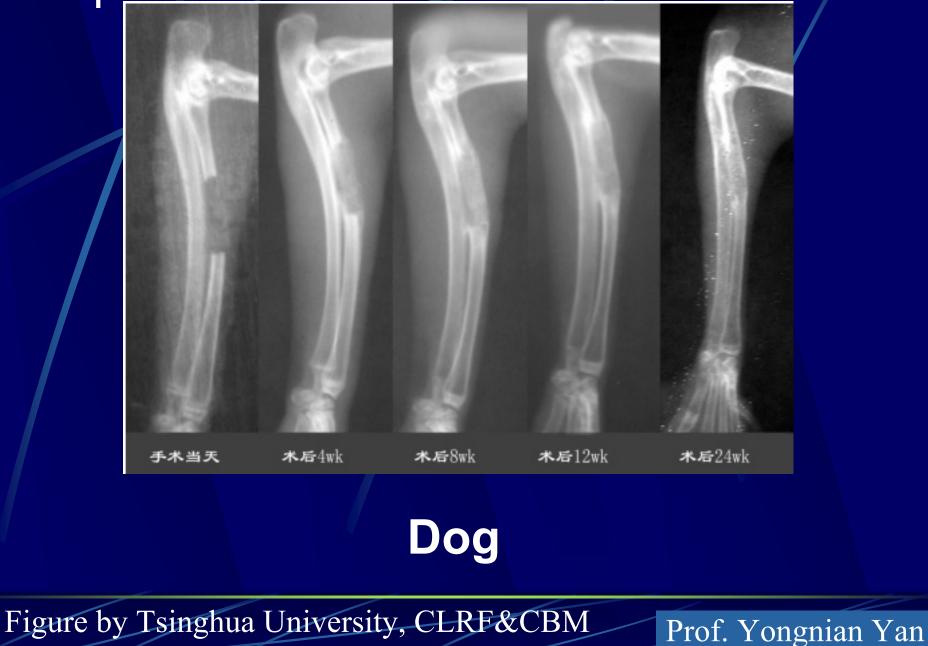
5 mm

15 mm

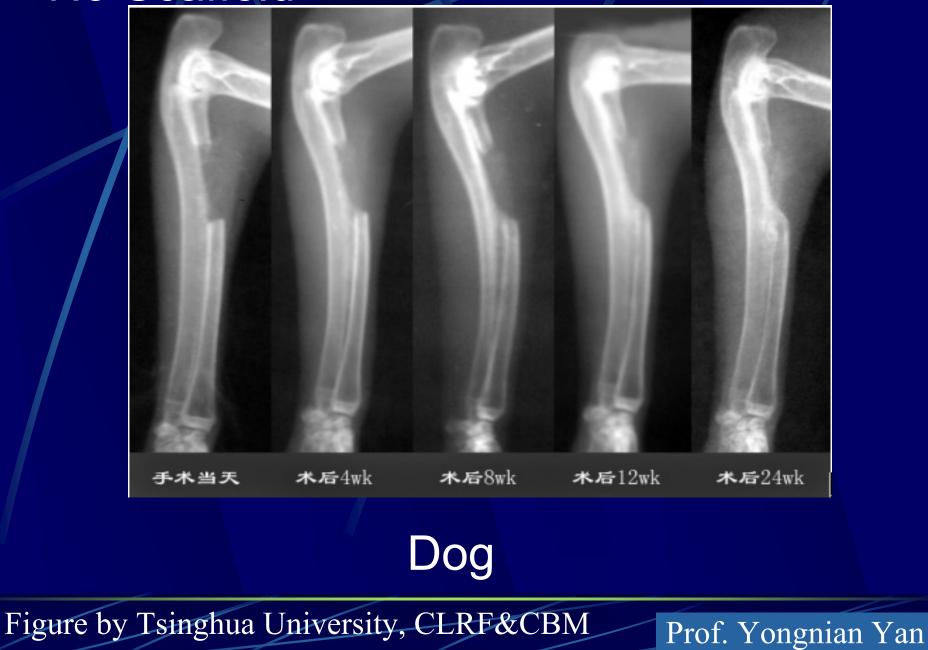
Figure by Tsinghua University, CLRF&CBM

10 mm

Implant bone Tissue Scaffold



No Scaffold



1. FFF Techuologies, FFF~Scaffold Mnuf. 2. Scaffold Manuf. Technologies 3. Non-degradation Scaffold **4.BONE Tissue Eng. Scaffolds** 5. 3-D cell Assembled 6. Laser Directed Guided Writing(LDGW) of cell

Prof. Yongnian Yan

Deeply integrating <u>manufacturing</u> <u>science</u> with <u>life science</u> and <u>cell</u> <u>biology</u>, viewing cells and the extracellular materials as assemble units

we propose the 3D controlled assembling by FFF to manufacture the Analogy Tissue Precursors (ATPs).

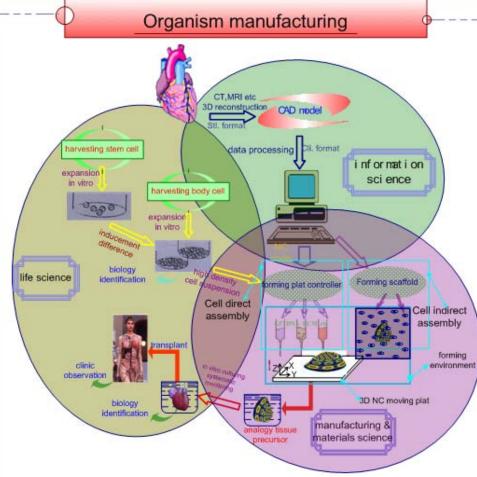


Analogy Tissue Precursor –

The 3D structure with the characteristic of living and metabolism



Illustration of the organism manufacturing



Two photos removed for copyright reasons.

Bioplotter

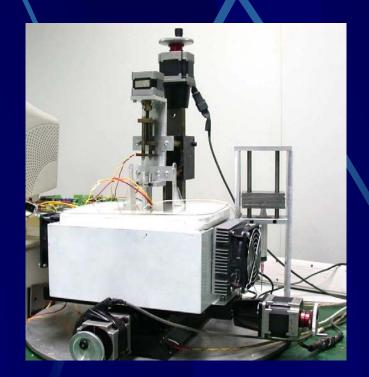
Landers&Mulhaupt(2000) EnvisionTec

Cell printer

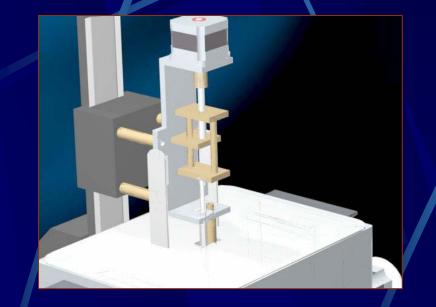
Vladimir Mironov, et al.,"Organ printing : computer-aided jet-based 3D tissue engineering",Biotechnology,Vol.21 No.4, April 2003



Cell controlled assembler



core parts of assembler



Odd nozzle extruding machine of Cell assembler



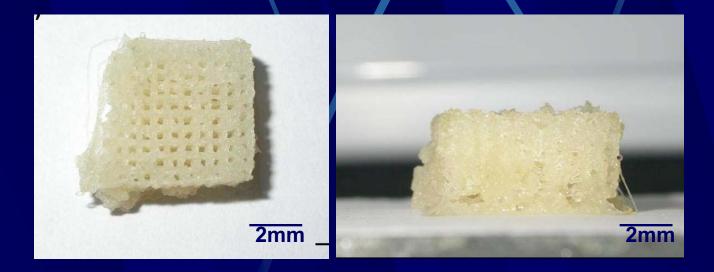
Multi-nozzle extruding machine of Cell assembler II

The table listed forming parameters

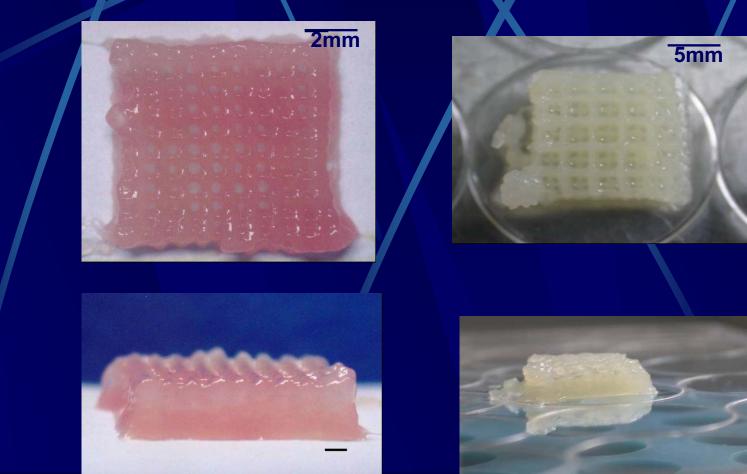
Extrusion cavity volume (ml)	1
Nozzle diameter (um)	200
Scanning speed (mm/s)	20
Extrusion frequency (Hz)	79
Material concentration (%)	5
Cross linker concentration	6
Lattice size (mm)	0.8

The experimented cells cartilage cell fibroblast cell hepatocyte cell endothelium cell myocardiac cell hepatocyte + fibroblast The experimented materials gelatin sodium alginate chitosan

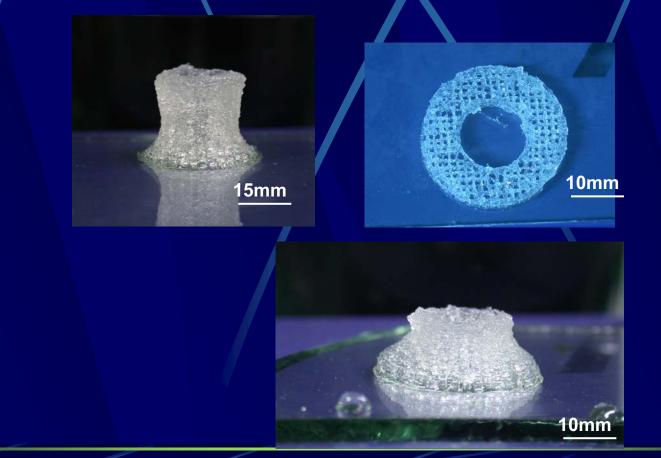
3D structure with hepatocyte/gelatin

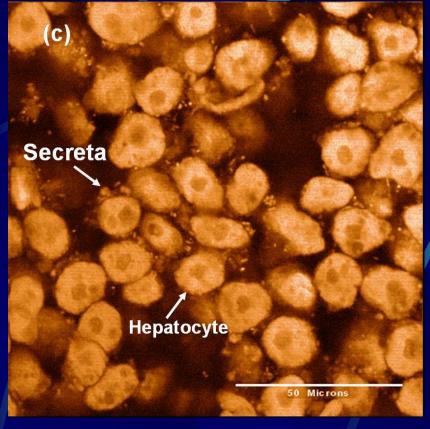


3D structure with hepatocyte/gelatin/sodium alginate



3D structure with chitosan





Confocal laser scanning (CLS) image of the hepatocytes One week after *in vitro* culture, stained with propidium iodide (PI,sigma USA)

Hepatic cells initially resided in the micro-environment provided by the 3D formed structure and presented large and round shape

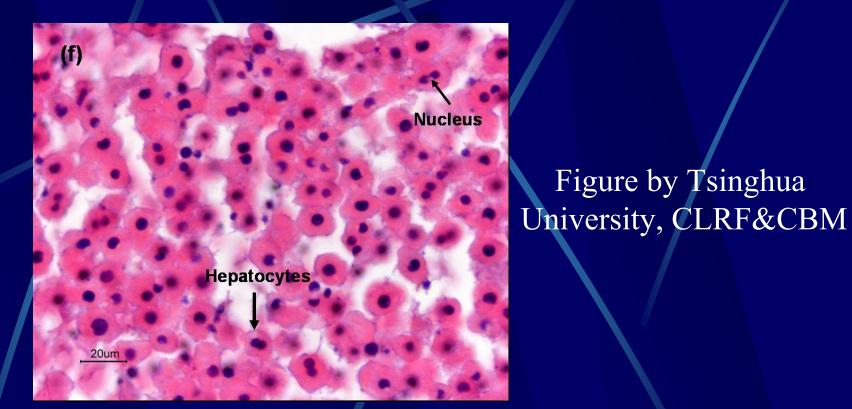


Image of histological section after two weeks in vitro, hepatic cells were still surviving and proliferating vigorously everywhere in the 3D structure, the long sinusoids were observed in many fields, as shown in picture.

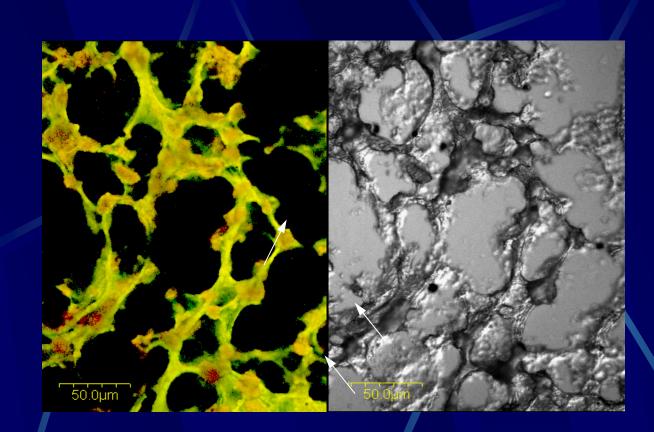
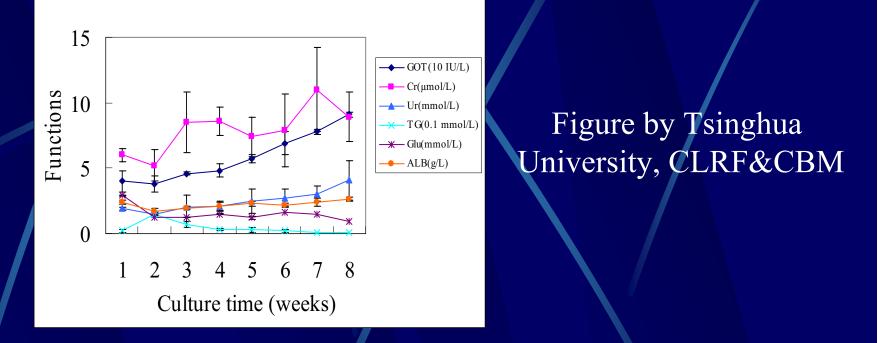


Figure by Tsinghua University, CLRF&CBM

LSC images of the hepatocytes after three weeks culture.
a) LSC observation with both PI staining and FITC-conjugation.
b) Negative control.
The cells displayed positive for albumin antigen-antibody reaction, and negative for the negative control of abnormal rabbit serum.
Arrows indicate the duct-like structures were formed.



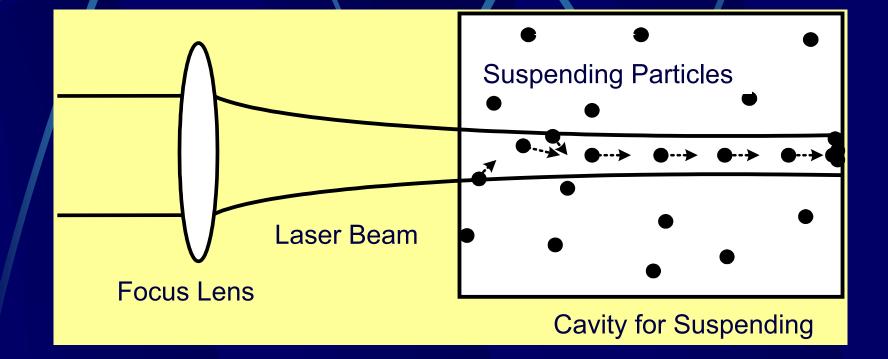
The amounts of albumin secretory and urea synthesis increase during 8 weeks culture. The amounts of albumin and the urea were in relative low level at the first 3 weeks, then increase in 3 to 6 weeks. After 6 weeks, the amounts kept in high level consistently. It indicates that hepatocytes perform liver-specific function in the network block.

Forming process:

1. FFF Technologies, FFF~Scaffold Mnuf. 2. Scaffold Manuf. Technologies 3. Non-degradation Scaffold **4.BONE Tissue Eng. Scaffolds** 5. 3-D cell Assembled 6. Laser Directed Guided Writing(LDGW) of cell



Principle of Laser Guided Direct Writing (LGDW) First posed by Renn, Michigan Institute of Technology, USA



Prof. Yongnian Yan

Influences of the medium on LGDW

1) flotage

(2) disturb of convection

(3) attenuation of the light power





Figure by Tsinghua University, CLRF&CBM

Prof. Yongnian Yan

Laser Power 500mW, girdling radius 15um Spacing between two dots 10~15um, Deposition time: 10min/dot

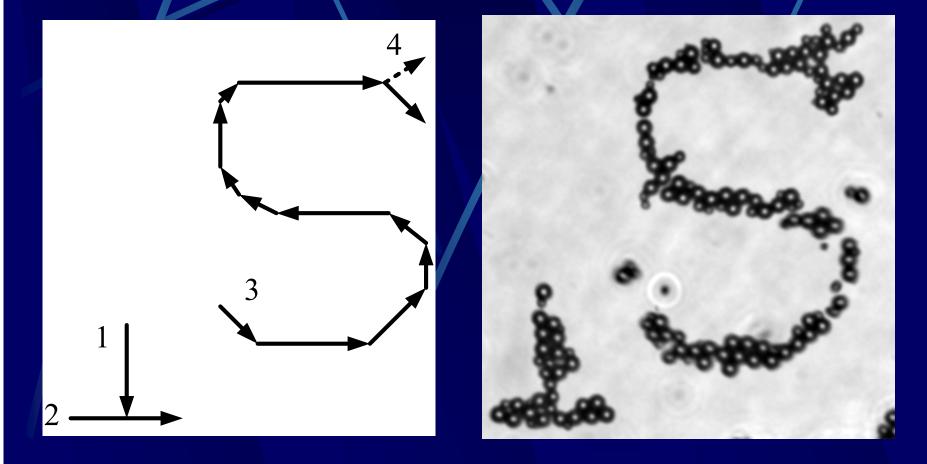


Figure by Tsinghua University, CLRF&CBM

Prof. Yongnian Yan